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# Potential Implications of the Phytohormone Abscisic Acid in Human Health Improvement at the Central Nervous System

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# Abstract

The plant hormone Abscisic Acid (ABA) has applications not only in agriculture, but also in human health. ABA is established as the key hormonal regulator of plant stress physiology, and it is also involved in plant growth and development under normal conditions. This phytohormone is present in the human body from dietary sources as well as from endogenous production through the carotenoid biogenesis pathway. ABA in mammals has both autocrine and paracrine function, and targets cells of the innate immune response, mesenchymal and hemopoietic stem cells and cells involved in the regulation of systemic glucose homeostasis, among others. Moreover, ABA increases glucose uptake in skeletal muscle and adipose tissue through an insulin-independent mechanism. Besides, ABA increases the energy expenditure in the brown and white adipose tissues. In this article, we review the potential of ABA to treat or ameliorate brain and spinal cord disorders, such as sleep disorders, depression, pain and Alzheimer derived memory impairments. Dietary ABA administration shows benefits in humans, as well as extensive data obtained in different mammal models and cell lines. Finally, future perspectives in nutraceutical use of ABA are discussed.

#### Introduction

Plants are central to our well-being, principally as food and medicine source. The coevolution between plants and humans is complex but long standing. Hominids have achieved different morphological and biochemical adaptations to plant material ingestion [1]. An interesting connection between plants and humans came from small signaling molecules called phytohormones [2]. Recent studies suggest that plant hormones also

work in mammalian systems, and have the potential to reduce human diseases such as cancer and diabetes [3]. In particular, we want to focus this review in the relationship between the phytohormone Abscisic Acid (ABA) and mammalian physiology at the central nervous system level. Extensive data obtained from people and different animal models and cell lines suggests a myriad of benefits of ABA in human health.



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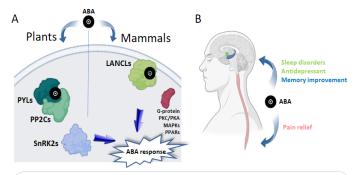
#### Abscisic acid machinery in plants

The phytohormone Abscisic Acid (ABA) is a key player in the plant stress response. The most studied role of ABA has been the induction of drought resistance, among other biotic and abiotic stresses, in both model plants and crops [4,5]. ABA also plays a role in different physiological processes, such as seed germination and early seedling growth, shoot and root growth and development, stomata closure, senescence, fruit ripening, fruit and leaf abscission, and bud dormancy [6].

In plants, abiotic stress (e.g., drought or salinity) induces ABA synthesis and active hormonal level increases notably. ABA synthesis in plants starts from the precursor carotene and follows the carotenoid pathway [7,8]. The initial steps, from carotene to xanthoxin are rendered in plastids and different enzymes are involved, for instance in *Arabidopsis thaliana*: viviparous (VPs), zeaxanthin epoxidase (ZEP/ABA1), abscisic acid (ABA)-deficient 4 (ABA4) and nine-cis-epoxycarotenoid dioxygenase (NCEDs). Then in the cytosol, ABA2 and ABA3 turn xanthoxin into active ABA. The rate-limiting step for ABA synthesis is the cleavage of 9-cis-epoxycarotenoid into xanthoxin by the NCEDs enzymes [9]. Moreover, transgenic plants with constitutive expression of NCEDs have high levels of ABA [10].

Net active ABA levels are set by the rate of synthesis and degradation/inactivation. In turn, ABA catabolism follows two different pathways: reversible conjugation or irreversible hydroxylation [7,11]. For instance, in *Arabidopsis thaliana* ABA can be glycosylated/inactivated by UGT71C5 to form ABA-glucose ester (ABA-GE), which is stored in vacuoles or in the endoplasmic reticulum. ABA-GE can be deconjugated to restore the active ABA by glycosidases, such as BG1 and BG2 [12-14]. This mechanism allows plants to promptly adapt to changes in the environment through ABA-mediated responses. By contrast, ABA can be irreversible converted to an inactive form as dihydrophaseic acid (DPA) by CYP707As and ABH2 [15,16].

In plants ABA is perceived inside the cell through the family of soluble receptors pyrabactin resistance 1 (PYR1)/PYR1-like (PYL)/regulatory components of ABA receptors (RCAR) [17-19] (Figure 1a). The PYL family has several members and has been identified in many crops, for instance 12 PYLs in palm [20], 14 PYLs in tomato [21], 20 PYLs in quinoa [22], 23 PYLs in benthamiana [23], 38 PYLs in wheat [24] and 46 PYLs in canola [25]. Upon ABA perception, the clade A protein phosphatases type 2Cs (PP2Cs) are inhibited through the formation of a ternary complex: ABA-PYL-PP2CA. This PP2CA inactivation relieves the



**Table 1:** ABA perception and its role in the central nervous system. A) ABA is perceived by the PYR/PYL receptors in plants and by the LANCL receptors in mammals. B) In the central nervous system ABA plays a role in sleep, depression, memory and pain. In green: hypothalamus. In blue: hippocampus. In red: spinal cord. Icons were obtained from Biorender.

inhibition of the ABA-activated subclass III SNF1-related protein kinases 2 (SnRK2s) [26,27]. Then, activated SnRK2 induces the activation of a battery of ABA effectors from transmembrane channels to transcription factors [28,29] (Figure 1a). In a second layer of regulation, the activity and half-life of components of this ABA core signaling pathway are regulated by different mechanisms such as the ubiquitin- 26S proteasome system, the endocytic/vacuolar degradation pathway, the circadian system and multiple secondary kinases [30-33].

# **ABA Signaling in Mammals**

One of the first reports describing the presence of ABA in mammals found the phytohormone in the central nervous system of pigs and rats [34]. The molecule purified from mammalian brain had the same biochemical properties than abscisic acid. Moreover, this brain factor inhibited stomatal apertures in Setcreasea pallida Rose (Commelinaceae) leaves. Later on, the endogenous synthesis of ABA by human granulocytes was also demonstrated [35]. Moreover, an increase of intracellular ABA levels after heat-stress (fever-like temperatures), and its release triggered by phagocytosis was also reported. For this reason ABA was proposed as a new endogenous pro-inflammatory cytokine in humans [35]. This idea of a phytohormone with a role in inflammation represents the first example of signaling module conservation, including the stress signal molecule and its transduction pathway, from plants to mammals. Moreover, this concept is also important from a clinical perspective, given that the identification of a new inflammation cytokine would magnify the possibility of development of new anti-inflammatory drugs (e.g. ABA antagonists molecules). Several other observations support the conclusion that ABA is endogenously produced by human and murine cells: granulocytes [35,36], macrophages and monocytes [37,38], insulin-releasing cells [39], mesenchymal stem cells (MSCs) [40], hemopoietic progenitors (HP) [41], adipocytes [42], keratinocytes [36], and fibroblasts [43], all have been shown to produce and release ABA when exposed to cell-specific stimuli.

Regarding ABA recognition in mammals, it was firstly reported that lanthionine synthetase C-like protein 2 (LANCL2) binds ABA and regulates cell glucose uptake and metabolism [44,45] (figure 1a). Later it was also shown that LANCL1 binds ABA, inducing the transcriptional expression of the glucose transporters GLUT4 and GLUT1 and the signaling proteins AMPK/PGC-1a/Sirt1, and also stimulates mitochondrial respiration and the expression of the skeletal muscle uncoupling proteins sarcolipin and UCP3 [46]. LANCL protein family, which includes LANCL1, 2 and 3, shares properties typical of a peptide and steroid hormone receptors. Both LANCL1 and LANCL2 bind ABA, although LANCL2 with higher affinity than LANCL1, with a Kd of 3 nM and 1  $\mu$ M, respectively [46,47]. Of note, none of the mammalian LANCL proteins are involved in lanthionine synthesis, even if the name includes "lanthionine synthetase" [48]. LANCL2 binds to the intracellular side of the plasma membrane through a myristoyl anchor [49], while LANCL1 and 3 are cytosolic soluble proteins [50]. On the other hand, LANCL1 and 2 are highly expressed in mammals, particularly in the brain, heart and germinal cells, with LANCL3 having the lowest expression levels of the LANCL proteins [46].

Upon ABA reception, LANCLs activates a G protein; in addition, as other steroid hormone receptors, they are capable of nuclear translocation after detachment from the membrane when de-myristoylated [46]. As proposed recently, this combination of peptide receptors (G protein coupling) and steroid hormone

receptors (with nuclear translocation), points to a heritage of the primordial origin of the hormone, or a consequence of ABA solubility properties [46]. In muscle cells, ABA recognition by LANCLs receptors lead to an insulin-independent glucose transport activation via the AMPK/PGC-1a pathway [51,52]. It was also reported that LANCL2 mediates akt activation via mTORC2 in human liver cells [53]. Besides, LANCL2 interacts with PPARy in white adipocytes, leading to PPARy-mediated activation of adipogenic genes after insulin-stimulated triglyceride accumulation [54]. In fact, LANCL2-\_/-\_ mice show a reduction in muscle activation and adipocyte glucose transport and metabolism, with the concomitant limitation in glucose tolerance [54]. Additionally, LANCL2 is also involved in the transcriptional activation of different browning genes in brown adipocytes [55]. The ABA signaling pathway in mammals also includes the cAMP-dependent activation of PKA and CD38 phosphorylation, leading to a cyclic ADP-ribose (cADPR)-mediated intracellular Ca<sup>2+</sup> increase [35,38,39,56].

#### **ABA Role in Brain Health**

#### ABA in sleep disorders

Sleep disorders increase health problems such as anxiety or forgetfulness [58,59]. Different therapies have been implemented to ameliorate insomnia, using GABA, melatonin and orexin receptors as pharmacological targets [60]. However, the drugs administered can cause dependency and other serious side effects [61,62]. An interesting alternative is the application of natural compounds such as ABA (Figure 1b).

It is well established that GABAergic neurotransmission has fundamental roles in boosting pentobarbital-induced sleep and relieving insomnia [63]. Indeed, ABA is involved in neurotransmitter release and regulates the activation of second messengers in both neural and non-neural cells [64-66]. On the other hand, the mammalian ABA receptor LANCL2 is related to plasma membrane and peroxisome proliferator activated receptors (PPARs), which are members of a nuclear hormone receptor superfamily with three subtypes (PPARα, PPARβ and PPARβ/δ) [44,67]. PPARs are located in different parts of the CNS, particularly in hypothalamic neurons [68] which are involved in sleep/ wake regulation [64]. Clinical and behavioral investigations have shown that PPARs have significant effects in sleep-wake cycle regulation [69]. Besides, it has been demonstrated that circadian locomotor activity is also affected by PPARs [70]. Sleep physiology and the circadian network are connected [71,72] and ABA-LANCL2-PPARy axis could be one of the links.

The efficacy of ABA to boost pentobarbital-induced sleep, and the involvement of GABA-A, PPARβ and PPARγ receptors in this process was recently demonstrated [73]. An ABA-induced promotion of sleep onset in rats was reported, with levels comparable to diazepam treatment [73]. On the other hand, it was reported that vitamin A plays a role in sleep cycle regulation and also has functional effects on the pineal gland [74,75]. ABA is a vitamin A-like lipophilic substance and has beneficial regulatory effects on brain physiology [76], with the potential to induce pro-hypnotic effects. Several reports show the ability of ABA to perform as a neuromodulator in the central nervous system, directly or indirectly interfering with synaptic neurotransmission due to changes in ion currents [65,66]. For instance, ABA interacts with neurotransmitters and second messengers such as glutamate, calcium and nitric oxide at synaptic levels [35,66,77]. The ABA hypnotic effect is probably induced by neurotransmitter regulation. However, this issue needs to be further investigated, and, above all, experiments in humans would be very welcome.

# ABA as an antidepressant

Following with the role of ABA in the central nervous system, some antidepressant effects of this phytohormone were also proposed [78,79] (Figure 1b). ABA is produced and released by the brain itself. Indeed, the brain contains much more ABA than any other type of tissue [34]; although with asymmetric distribution, and the hypothalamus showed the highest ABA concentration. Depression is a stress-related disorder and affects more than 10% of the world population [80,81]. The stress response in mammals is mainly regulated by the hypothalamus and a dysfunction in the hypothalamic-pituitary-adrenal (HPA) axis is usually involved in depression symptoms [82,83]. The ABA abundance in the hypothalamus suggests a role of ABA in the stress and depression response.

An association between retinoic acid (RA) and depressive symptoms has also been reported [84,85]. For instance, RA chronic administration induces HPA axis hyperactivity and depression-like behavioral changes in rats [86]. In addition, depressed patients exhibit a dysregulation in brain retinoid [87,88]. Interestingly, ABA and RA are carotenoid derivatives [7,89], and both molecules share a similar structure, specially a key carboxyl group in the isoprene-composed side chain involved in their bioactivity [84,90].

The corticotrophin-Releasing Hormone (CRH) in the paraventricular nucleus of the hypothalamus plays a central role in the regulation of HPA axis activity [82,83]. The release of corticosterone and stimulus-induced c-fos expression are widely used as markers in studies of neuronal activation after stress [91,92]. Interestingly, the ABA concentration significantly increased in the serum after stress treatments, and correlates with elevated corticosterone and c-fos levels [78]. By contrast, a decrease in ABA concentration was found in the hypothalamus of the rats under acute stress. These results are suggesting ABA may play a role in the stress response. Indeed, chronic ABA administration in rats showed a downregulation in CRH mRNA expression in the hypothalamus. Moreover, lower corticosterone concentrations in the serum were found after ABA treatment. These results indicate that ABA inhibits the HPA axis activity under physiological conditions.

On the other hand, chronic ABA treatment induces sucrose intake in rats. Sucrose intake correlates with the motivation to seek out a pleasurable experience, and this is connected with the capacity to feel interest or pleasure in mammals [93]. The ABA- induced higher sucrose intake and the downregulated HPA axis activity suggest that this phytohormone may play a role in the pathogenesis of depression. Indeed, the antidepressant effect of ABA was recently demonstrated in rats and mice under Chronic Unpredictable Mild Stress (CUMS) and Forced Swimming Test (FST) [78,79]. CUMS successfully decreased sucrose intake and increased immobility in the FST in rats, while ABA improved these depression-like behaviors [78]. Anhedonia is another behavior related with depression symptoms and sucrose intake [93,94]. ABA-treated rats spent a longer time swimming in the FST compared with the CUMS rats, although this accordance could not alleviate anxiety-related behaviors [78]. In agreement, ABA induces the normalization of CRH expression in the hypothalamus to control levels, and decreases corticosterone levels in serum. These results demonstrate the anti-depressant activity of ABA at the central nervous system,

and underscore the potential of this phytohormone for novel therapeutic strategies development to treat depression.

# **ABA** improves memory in mammals

An interesting ABA function described in mammals is the positive effect that this phytohormone plays on spatial learning and memory performance [64,66] (Figure 1b). Indeed, this role of ABA is also involved in the amelioration of cognitive impairment in diseases such as obesity induced type 2 diabetes [95,96], Alzheimer disease [97-99] and essential tremor [100]. ABA not only readily permeates the brain when applied peripherally [64], but is also produced and released by the brain itself [34,56]. Moreover, the high ABA levels seen in the hippocampus suggest a connection between this phytohormone and learning/memory processes. This is also supported by the fact that ABA and Retinoic Acid (RA) share similar molecular structures, and RA has been reported to improve spatial memory in rodents [101-103]. Indeed, ABA has a positive effect on spatial learning and memory performance [66]. Furthermore, the PI3K/PKC signaling pathway is involved in this mechanism given that its inhibitors suppress the ABA-induced learning and memory improvement [66]. The serine/threonine kinase PKC also participates in memory related disorders such as Alzheimer's disease [104].

Alzheimer's disease is a type of dementia related to neurodegenerative processes in the elderly,; and neuroinflammation is one of the most important pathological causes [105,106]. ABA treatment improves memory impairment in Alzheimer's disease 5xFAD model mice, through neuroinflammation inhibition and LANCL2/CREB upregulation in the cortex and hippocampus [97]. Furthermore, this role of the phytohormone was also observed in the triple transgenic mice (3xTg-AD), another murine model of Alzheimer's disease [99]. Even more, ABA also ameliorates cognitive impairments in a streptozotocin-induced rat model of Alzheimer's disease [98]. Streptozotocin central injection produces neuroinflammation and oxidative stress in the brain, leading to learning and memory impairments [107], and ABA administration attenuates these deficits through activation of PPAR $\beta/\delta$  and PKA signaling [98]. These results together illustrate that ABA is an effective treatment to improve cognitive health.

#### Role of ABA in pain treatment

It was recently demonstrated that ABA elicits antinociceptive effects and reduces neuropathic pain at spinal cord level [108-110] (Figure 1b). The spinal cord plays a key role in pain transmission, regulation and processing. In particular, the dorsal horn parts and laminas have a paramount importance in pain control and transmission [111]. Injury and dysfunctionalrelated neuropathic pain treatment in the nervous system represents a current clinical challenge given the relative lack of potent and safe analgesics [112]. Neuropathic pain originates from an aberrant neuronal activity along the pain signaling pathway, and neurons in the spinal dorsal horn are involved in this process [113,114]. Furthermore, neuroinflammation in the spinal dorsal horn is a prerequisite for a dysfunction of spinal neuronal activation and the genesis of neuropathic pain [115-117]. Moreover, neuroinflammation involves leukocytes infiltration, microglia and astrocytes activation, and pro-inflammatory cytokines over-production [115-116]. As already stated, identifying signaling molecules controlling neuroinflammation would provide novel molecular targets for the development of novel analgesics [108].

The presence of ABA in the spinal dorsal horn was recently reported [108] ABA concentrations in this tissue are not significantly altered by peripheral nerve injury- induced neuroinflammation. Moreover, ABA treatment ameliorates spinal inflammation and chronic pain in rats [108]. On the other hand, it was shown that the mammalian ABA receptor LANCL2 is expressed in immune cells such as T cells, macrophages, dendritic cells and spinal microglia [61,108]. Furthermore, knockdown of the LANCL2 gene with siRNA in the spinal dorsal horn recapitulates the nerve injury induced spinal neuroinflammation and nociceptive behaviors [108]. In addition, spinal microglia cells respond to ABA treatment [56]. Indeed, the abundance of LANCL2 was reduced in the spinal cord with nerve injury- induced neuroinflammation, and this reduction was reverted by ABA treatment [108]. Moreover, ABA treatment prevented the reduction in LANCL2 protein expression in the cortex of an Alzheimer's disease mouse model [97].

The role of the ABA and LANCL2 signaling on mammalian inflammatory signaling pathways is controversial, given that both pro-inflammatory and anti-inflammatory effects of this axis have been reported [35,36,95,97,109,118]. Possibly, the ABA/LANCL2 function on inflammation is tissue/organ-specific. For instance, pro-inflammatory activity was reported in ABA treated granulocytes that showed an increment in phagocytosis, production of reactive oxygen species (ROS) and nitric oxide (NO) [35]. In accordance, an increment of TNFα, NO, and ROS dependent of ultraviolet light induced ABA release from granulocytes and keratinocytes has been demonstrated [36]. On the contrary, an anti-inflammatory ABA activity was induced in animals with inflammatory bowel disease, since this treatment reduces TNFα expression and macrophage infiltration in white adipose tissue [118]. Besides, an ABA- induced reduction of glial activation and production of TNFα and IL-1β was reported, leading to an improvement of cognitive function in the brain of a murine model of Alzheimer's disease [97]. In this sense, ABA treatment also reduces high fat diet- induced microglial activation and TNF $\alpha$  production in the hypothalamus of rats [95]. Moreover, ABA treatment attenuates spinal neuroinflammation induced by nerve injury in the spinal cord, and reduces Iba1 and TNF $\alpha$  expression [108]. An explanation proposed elsewhere, is that two different signaling pathways may be working for the opposite inflammatory ABA induced responses [119]. Supporting this idea, it was shown that ABA- induced pro-inflammatory responses in granulocytes is mediated by a pertussis toxin (PTX)-sensitive G-protein [35]. In contrast, ABA- induced anti-inflammatory activity is not affected by PTX, given that ABA treatment still attenuated lipopolysaccharide- induced microglial activation and TNFα production in the spinal cord, suggesting that G-protein is dispensable in this scenario [108].

In another line of evidence, intrathecal ABA administration in rats lead to analgesia in tail-flick and hot-plate tests [110]. Furthermore, intracerebroventricular ABA application showed a potent pain-relieving activity in rats under formalin tests [109]. The molecular mechanism of these phenotypes is PKA-dependent and involves p-ERK down-regulation, as well as the peroxisome proliferator-activated receptors (PPAR  $\beta/\delta$ ) and opioid signaling activation [109-110]. Interestingly, opioids induce antinociception via PTX- sensitive inhibitory G-proteins [120]. Of note, ABA is structurally similar to the PPARy agonist thiazolidinediones and both compounds ameliorate insulin resistance and inhibit systemic inflammation [67,121]. Besides, PPAR receptors are members of the nuclear receptor superfamily [122].

ABA plays a critical role in the genesis of neuropathic pain and showed antinociceptive effects [108-110]. The fact that deficiencies in ABA reception and signaling in mammals can be remedied by exogenous ABA application provides a rationale to explore neuropathic pain treatments with this phytohormone. Moreover, given that ABA is present in a vegetable and fruit-based diet, it is also conceivable to explore the nutraceutical application of ABA in the neuropathic pain field.

#### Conclusion and future perspectives

There is substantial evidence to argue that ABA plays a neurotrophic role in the mammalian central nervous system, related with sleep, depression, pain and memory (figure 1b). While these evidences are mostly based on animal models and cell lines, further insight into ABA functions in the human brain would be necessary in order to determine its potential therapeutic effect.

Despite the fact that ABA is produced and released by the brain itself [34], it is also conceivable to study the nutraceutical application of this phytohormone given that it readily permeates the blood brain barrier [64]. A fruit and vegetable rich food diet represents a natural source of ABA [123]. In particular avocados (2.0 mg/kg), citrus (1.25 mg/kg), soybean (0.79 mg/kg) and figs (0.72 mg/kg) contain high ABA levels [123]. An interesting field of research would be to generate crops with magnified ABA levels through abiotic stress treatments or genome editing. Alternatively, the feasibility of ABA production in bioreactors was recently demonstrated, using the oleaginous yeast *Yarrowia lipolytica* [124].

Finally, an increasingly used tool in agriculture is the use of ABA agonists to combat the severe drought episodes induced by the climate change [125-126]. Some of these agonists are even more potent and persistent than ABA in crops. Studies using ABA agonists in mammals have not been reported yet, representing a long and promising road ahead in this field.

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#### References

- 1. Barbara S. Plants and people: Our shared history and future. Plants people planet 2019; 1: 14-19.
- 2. Davies PJ. The Plant Hormones: Their Nature, Occurrence, and Functions. Book Chapter in: Plant Hormones. 2010
- Mukherjee A, Gaurav AK, Singh S, Yadav S, Bhowmick S, et al. The bioactive potential of phytohormones: A review. Biotechnol Rep. 2022; 35: e00748.
- Chen K, Li G, Bressan RA, Song C, Zhu J, et al. Abscisic acid dynamics, signaling, and functions in plants. J ntegr Plant Biol. 2020; 62: 25-54.
- Ruiz-Partida R, Rosario S, Lozano-Juste J. An Update on Crop ABA Receptors. Plants. 2021; 10: 1087.
- 6. Cutler SR, Rodriguez PL, Finkelstein RR, Abrams SR. Abscisic Acid: Emergence of a Core Signaling Network. Annu Rev Plant Biol. 2010; 61: 651-679.
- Nambara E, Marion-Poll A. Abscisic acid biosynthesis and catabolism. Annu Rev Plant Biol. 2005; 56: 165-185.

- Arc E, Sechet J, Corbineau F, Rajjou L, Marion-Poll A. ABA crosstalk with ethylene and nitric oxide in seed dormancy and germination. Front Plant Sci. 2013; 4: 63.
- Qin X, Zeevaart JAD. The 9-cis-epoxycarotenoid cleavage reaction is the key regulatory step of abscisic acid biosynthesis in water-stressed bean. Proc Natl Acad Sci. USA 1999; 96: 15354-15361.
- Qin X, Zeevaart JAD. Overexpression of a 9-cis-Epoxycarotenoid Dioxygenase Gene in Nicotiana plumbaginifolia Increases Abscisic Acid and Phaseic Acid Levels and Enhances Drought Tolerance. Plant Physiol. 2002; 128: 544-551.
- Ali F, Qanmber G, Li F, Wang Z. Updated role of ABA in seed maturation, dormancy, and germination. J Adv Res. 2021; 35:199-214.
- Lee KH, Piao HL, Kim HY, Choi SM, Jiang F, et al. Activation of Glucosidase via Stress-Induced Polymerization Rapidly Increases Active Pools of Abscisic Acid. Cell. 2006; 126: 1109-1120.
- Xu ZY, Lee KH, Dong T, Jeong JC, Jin JB, et al. A vacuolar betaglucosidase homolog that possesses glucose-conjugated abscisic acid hydrolyzing activity plays an important role in osmotic stress responses in Arabidopsis. Plant Cell. 2012; 24: 2184-2199.
- 14. Liu Z, Yan JP, Li DK, Luo Q, Yan Q, et al. UDP-Glucosyltransferase71C5, a Major Glucosyltransferase, Mediates Abscisic Acid Homeostasis in Arabidopsis. Plant Physiol. 2015; 167: 1659-1670.
- Kushiro T, Okamoto M, Nakabayashi K, Yamagishi K, Kitamura S, et al. The Arabidopsis cytochrome P450 CYP707A encodes ABA 80-hydroxylases: Key enzymes in ABA catabolism. EMBO J. 2004; 23: 1647-1656.
- Weng JK, Ye M, Li B, Noel JP. Co-evolution of Hormone Metabolism and Signaling Networks Expands Plant Adaptive Plasticity. Cell. 2016; 166: 881-893.
- Park SY, Fung P, Nishimura N, Jensen DR, Fujii H, et al. Abscisic Acid Inhibits Type 2C Protein Phosphatases via the PYR/PYL Family of START Proteins. Science. 2009; 324: 1068-1071.
- Ma Y, Szostkiewicz I, Korte A, Moes D, Yang Y, et al. Regulators of PP2C Phosphatase Activity Function as Abscisic Acid Sensors. Science. 2009; 324: 1064-1068.
- Santiago J, Dupeux F, Round A, Antoni R, Park SY, et al. The abscisic acid receptor PYR1 in complex with abscisic acid. Nature 2009; 462: 665-668.
- Garcia-Maquilon I, Coego A, Lozano-Juste J, Messerer M, de Ollas C, et al. PYL8 ABA receptors of Phoenix dactylifera play a crucial role in response to abiotic stress and are stabilized by ABA. J Exp Bot. 2021; 72: 757-774.
- Sun L, Wang YP, Chen P, Ren J, Ji K, et al. Transcriptional regulation of SIPYL, SIPP2C, and SISnRK2 gene families encoding ABA signal core components during tomato fruit development and drought stress. J Exp Bot. 2011; 15: 5659-5669.
- Pizzio GA. Genome-Wide Identification of the PYL Gene Family in Chenopodium quinoa: From Genes to Protein 3D Structure Analysis. Stresses. 2022; 2: 290-307
- Pizzio GA, Mayordomo C, Lozano-Juste J, Garcia-Carpintero V, Vazquez-Vilar M, et al. PYL1- and PYL8-like ABA Receptors of Nicotiana benthamiana Play a Key Role in ABA Response in Seed and Vegetative Tissue. Cells. 2022; 11: 795.
- 24. Lei P, Wei X, Gao R, Huo F, Nie X, et al. Genome-wide identification of PYL gene family in wheat: Evolution, expression and 3D structure analysis. Genomics. 2021; 113: 854-866.

- Di FF, Jian HJ, Wang TY, Chen XP, Ding YR, et al. Genome-wide analysis of the PYL gene family and identification of PYL genes that respond to abiotic stress in Brassica napus. Genes. 2018; 9: 156.
- Umezawa T, Sugiyama N, Mizoguchi M, Hayashi S, Myouga F, et al. Type 2C protein phosphatases directly regulate abscisic acidactivated protein kinases in Arabidopsis. Proc Natl Acad Sci USA 2009; 106: 17588-17593.
- Vlad F, Rubio S, Rodrigues A, Sirichandra C, Belin C, et al. Protein Phosphatases 2C Regulate the Activation of the Snf1-Related Kinase OST1 by Abscisic Acid in Arabidopsis. Plant Cell. 2009; 21: 3170-3184.
- Huang D, Wu W, Abrams SR, Cutler AJ. The relationship of drought-related gene expression in Arabidopsis thaliana to hormonal and environmental factors. J Exp Bot. 2008; 59: 2991-3007
- Fujita Y, Fujita M, Shinozaki K, Yamaguchi-Shinozaki K. ABA-mediated transcriptional regulation in response to osmotic stress in plants. J Plant Res. 2011; 124: 509-525
- 30. Zhu JK. Abiotic stress signaling and responses in plants. Cell. 2016; 167: 313-324.
- Coego A, Julian J, Lozano-Juste J, Pizzio GA, Alrefaei A, et al. Ubiquitylation of ABA Receptors and Protein Phosphatase 2C Coreceptors to Modulate ABA Signaling and Stress Response. Int J Mol Sci. 2021; 22: 7103.
- 32. Pizzio GA. Abscisic Acid Machinery Is under Circadian Clock Regulation at Multiple Levels. Stresses. 2022; 2: 65-78.
- 33. Pizzio GA, Rodriguez PL. Dual regulation of SnRK2 signaling by Raf-like MAPKKKs. Molecular Plant. 2022; 15: 1260-1262
- Le Page-Degivry MT, Bidard JN, Rouvier E, Bulard C, Lazdunski M.
  Presence of abscisic acid, a phytohormone, in the mammalian brain. Proc Nati Acad Sci USA. 1986; 83: 1155-1158,
- 35. Bruzzone S, Moreschi I, Usai C, Guida L, Damonte G, et al. Abscisic acid is an endogenous cytokine in human granulocytes with cyclic ADP- ribose as second messenger. Proc Natl Acad Sci U S A. 2007; 104: 5759-5764.
- Bruzzone S, Basile G, Mannino E, Sturla L, Magnone M, et al. Autocrine abscisic acid mediates the UV-B-induced inflammatory response in human granulocytes and keratinocytes. J Cell Physiol. 2012; 227: 2502-2510.
- Magnone M, Sturla L, Jacchetti E, Scarfi S, Bruzzone S, et al. Autocrine abscisic acid plays a key role in quartz-induced macrophage activation. FASEB J. 2012; 26: 1261-1271.
- 38. Magnone M, Bruzzone S, Guida L, Damonte G, Millo E, et al. Abscisic acid released by human monocytes activates monocytes and vascular smooth muscle cell responses involved in atherogenesis. J Biol Chem. 2009; 284: 17808-17818.
- Bruzzone S, Bodrato N, Usai C, Guida L, Moreschi I, et al. Abscisic acid is an endogenous stimulator of insulin release from human pancreatic islets with cyclic ADP ribose as second messenger. J. Biol. Chem. 2008; 283: 32188-321897.
- 40. Scarfi S, Ferraris C, Fruscione F, Fresia C, Guida L, et al. Cyclic ADP-ribose-mediated expansion and stimulation of human mesenchymal stem cells by the plant hormone abscisic acid. Stem Cells. 2008; 26: 2855-2864.
- 41. Scarfi S, Fresia C, Ferraris C, Bruzzone S, Fruscione F, et al. The plant hormone abscisic acid stimulates the proliferation of human hemopoietic progenitors through the second messenger cyclic ADP-ribose. Stem Cells. 2009; 27: 2469-2477.

- 42. Bruzzone S, Ameri P, Briatore L, Mannino E, Basile G, et al. The plant hormone abscisic acid increases in human plasma after hyperglycemia and stimulates glucose consumption by adipocytes and myoblasts. FASEB J. 2012; 26: 1251-1260.
- Bruzzone S, Battaglia F, Mannino E, Parodi A, Fruscione F, et al. Abscisic acid ameliorates the systemic sclerosis fibroblast phenotype in vitro. Biochem. Biophys. Res Commun. 2012; 422: 70-74
- Sturla L, Fresia C, Guida L, Bruzzone S, Scarfi S, et al. LANCL2 is necessary for abscisic acid binding and signaling in human granulocytes and in rat insulinoma cells. J Biol Chem. 2009; 284: 28045e28057.
- Sturla L, Fresia C, Guida L, Grozio A, Vigliarolo T, et al. Binding of abscisic acid to human LANCL2. Biochem Biophys Res Commun. 2011; 415: 390-395.
- 46. Spinelli S, Begani G, Guida L, Magnone M, Galante D, et al. LANCL1 binds abscisic acid and stimulates glucose transport and mitochondrial respiration in muscle cells via the AMPK/PGC- $1\alpha$ / Sirt1 pathway. Mol Metab. 2021; 53: 101263.
- 47. Cichero E, Fresia C, Guida L, Booz V, Millo E, et al. Identification of a high affinity binding site for abscisic acid on human lanthionine synthetase component C-like protein 2. Int. J. BioChem. Cell Biol. 2018; 97: 52-61.
- He C, Zeng M, Dutta D, Koh TH, Chen J, et al. LanCL proteins are not involved in lanthionine synthesis in mammals. Sci Rep. 2017; 7: 40980.
- 49. Landlinger C, Salzer U, Prohaska R. Myristoylation of human LanClike protein 2 (LANCL2) is essential for the interaction with the plasma membrane and the increase in cellular sensitivity to Adriamycin. Biochimica et Biophysica Acta. 2006; 1758: 1759e1767.
- 50. Fresia C, Vigliarolo T, Guida L, Booz V, Bruzzone S, et al. G-protein coupling and nuclear translocation of the human abscisic acid receptor LANCL2. Sci Rep. 2016; 6: 26658.
- Magnone M, Sturla L, Guida L, Spinelli S, Begani G, et al. Abscisic Acid: A Conserved Hormone in Plants and Humans and a Promising Aid to Combat Prediabetes and the Metabolic Syndrome. Nutrients. 2020; 12: 1724.
- 52. Puce S, Basile G, Bavestrello G, Bruzzone S, Cerrano C, et al. Abscisic acid signaling through cyclic ADP-ribose in hydroid regeneration. J Biol Chem. 2004; 279: 39783-39788.
- Zeng M, Van der Donk WA, Chen J. Lanthionine synthetase C-like protein 2 (LanCL2) is a novel regulator of Akt. Mo Biol Cell 2014; 25: 3954e3961.
- 54. Dutta D, Lai KY, Reyes-Ordoñez A, Chen J, Van der Donk WA. Lanthionine synthetase C-like protein 2 (LanCL2) is important for adipogenic differentiation. J Lipid Res. 2018; 59: 1433e1445.
- Leckie CP, McAinsh MR, Allen GJ, Sanders D, Hetherington AM.
  Abscisic acid-induced stomatal closure mediated by cyclic ADPribose. Proc Natl Acad Sci USA. 1998; 95: 15837-15842.
- Bodrato N, Franco L, Fresia C, Guida L, Usai C, et al. Abscisic acid activates the murine microglial cell line N9 through the second messenger cyclic ADP-ribose. J Biol Chem. 2009; 284: 14777-14787.
- Bodrato N, Franco L, Fresia C, Guida L, Usai C, et al. Abscisic acid activates the murine microglial cell line N9 through the second messenger cyclic ADP-ribose. J Biol Chem. 2009; 284: 14777-14787

- Li Y, Zhang S, Zhu J, Du X, Huang F. Sleep disturbances are associated with increased pain, disease activity, depression, and anxiety in ankylosing spondylitis: a case-control study. Arthritis Res Ther. 2012; 14: R215.
- Chokroverty S. Overview of sleep & sleep disorders. Indian J Med Res. 2010; 131: 126-140.
- Rihel J, Schier AF. Sites of action of sleep and wake drugs: insights from model organisms. Curr Opin Neurobiol. 2013; 23: 831-840.
- 61. Griffiths AN, Jones DM, Richens A. Zopiclone produces effects on human performance similar to flurazepam, lormetazepam and triazolam. Br Clin Pharmacol. 1986; 21: 647-653.
- 62. Bocca ML, Le Doze F, Etard O, Pottier M, L'Hoste J, et al. Residual effect of zolpidem 10 mg and zopiclone 7.5 mg versus flunitrazepam 1 mg and placebo on driving performance and ocularsaccades. Psychopharmacology. 1999; 143: 373-379.
- Liao JF, Huang SY, Jan YM, Yu LL, Chen CF. Central inhibitory effects of water extract of Acori graminei rhizoma in mice. J Ethnopharmacol. 1998; 61: 185-193.
- 64. Qi CC, Ge JF, Zhou JN. Preliminary evidence that abscisic acid improves spatial memory in rats. Physiol Behav. 2015; 139: 231-239.
- Li HH, Hao RL, Wu SS, Guo PC, Chen CJ, et al. Occurrence, function and potential medicinal applications of the phytohormone abscisic acid in animals and humans. Biochem Pharmacol. 2011; 82: 701-712.
- 66. Naderi R, Esmaeili-Mahani S, Abbasnejad M. Phosphatidylinositol- 3-kinase and protein kinase C are involved in the pro-cognitive and anti-anxiety effects of phytohormone abscisic acid in rats. Biomed Pharmacother. 2017; 96: 112-119.
- 67. Guri AJ, Evans NP, Hontecillas R, Bassaganya-Riera J. T cell PPAR γ is required for the anti-inflammatory efficacy of abscisic acid against experimental IBD. J Nutr Bioch. 2011; 22: 812-819.
- 68. Moreno S, Farioli-Vecchioli S, Cerù MP. Immunolocalization of peroxisome proliferator-activated receptors and retinoid X receptors in the adult rat CNS. Neurosci. 2004; 123: 131-45.
- 69. Mijangos-Moreno S, Poot-Aké A, Guzmán K, Arankowsky-Sandoval G, Arias-Carrión O, et al. Sleep and neurochemical modulation by the nuclear peroxisome proliferator-activated receptor  $\alpha$  (PPAR- $\alpha$ ) in rat. Neurosci Res. 2016; 105: 65-69.
- Shirai H, Oishi K, Kudo T, Shibata S, Ishida N. PPARα is a potential therapeutic target of drugs to treat circadian rhythm sleep disorders. Bioch Biophys Res Commun. 2007; 357: 679-682.
- 71. Oishi K, Atsumi GI, Sugiyama S, Kodomari I, Kasamatsu M, et al. Disrupted fat absorption attenuates obesity induced by a high-fat diet in Clock mutant mice. FEBS Lett. 2006; 580: 127-130.
- Zvonic S, Floyd ZE, Mynatt RL, Gimble JM. Circadian rhythms and the regulation of metabolic tissue function and energy homeostasis. Obesity (Silver Spring). 2007; 15: 539-543.
- Madadzadeh M, Abbasnejad M, Mollashahi M, Pourrahimi AM, Esmaeili-Mahani S. Phytohormone abscisic acid boosts pentobarbital-induced sleep through activation of GABA-A, PPARβ and PPARγ receptor signaling. Arq. Neuropsiquiatr. 2021; 79: 216-221.
- 74. Sei H. Vitamin A and sleep regulation. J Med Invest. 2008; 55(1-2): 1-8.
- 75. Guillaumond F, Giraudet F, Becquet D, Sage D, Laforge-Anglade G, et al. Vitamin A is a necessary factor for sympatheticindependent rhythmic activation of mitogen-activated protein kinase in the rat pineal gland. Eur J Neurosci. 2005; 21: 798-802.

- 76. Finkelstein RR, Gampala SS, Rock CD. Abscisic acid signaling in seeds and seedlings. Plant Cell. 2002; 14: S15-45
- Pidoplichko VI, Reymann KG. Abscisic acid potentiates NMDAgated currents in hippocampal neurones. Neuroreport. 1994; 5: 2311-2316.
- Qi CC, Zhang Z, Fang H, Liu J, Zhou N, et al. Antidepressant effects of abscisic acid mediated by the downregulation of corticotrophin-releasing hormone gene expression in rats. Int J Neuropsychopharmacol. 2014; 18: pyu006.
- Qi CC, Shu YM, Chen FH, Ding YQ, Zhou JN. Sensitivity during the forced swim test is a key factor in evaluating the antidepressant effects of abscisic acid in mice. Behav Brain Res. 2016; 300: 106-113.
- Lu M, Yang JZ, Geng F, Ding JH, Hu G. Iptakalim confers an antidepressant effect in a chronic mild stress model of depression through regulating neuro-inflammation and neurogenesis. Int J Neuropsychopharmacol. 2014; 7: 1501-1510.
- Chen Y, Jiang T, Chen P, Ouyang J, Xu G, et al. Emerging tendency towards autoimmune process in major depressive patients: a novel insight from Th17 cells. Psychiatry Res. 2011; 188: 224-230.
- Swaab DF. Development of the human hypothalamus. Neurochem Res. 1995; 20: 509-519.
- 83. Bao AM, Fischer DF, Wu YH, Hol EM, Balesar R, et al. A direct androgenic involvement in the expression of human corticotropin-releasing hormone. Mol. Psychiatr. 2006; 11: 567-576.
- 84. Bremner JD, McCaffery P. The neurobiology of retinoic acid in affective disorders. Prog. Neuropsychopharmacol. Biol Psychiat. 2008; 32: 315-331.
- Bremner JD, Shearer KD, McCaffery PJ. Retinoic acid and affective disorders: the evidence for an association. J Clin Psychiat. 2012; 73: 37-50.
- Cai L, Yan XB, Chen XN, Meng QY, Zhou JN. Chronic alltrans retinoic acid administration induced hyperactivity of HPA axis and behavioral changes in young rats. Eur. Neuropsychopharmacol. 2010; 20: 839-847.
- 87. Chen XN, Meng QY, Bao AM, Swaab DF, Wang GH, et al. The involvement of retinoic acid receptor-alpha in corticotropin-releasing hormone gene expression and affective disorders. Biol Psychiat. 2009; 66: 832-839.
- 88. Qi XR, Kamphuis W, Wang S, Wang Q, Lucassen PJ, et al. Aberrant stress hormone receptor balance in the human prefrontal cortex and hypothalamic paraventricular nucleus of depressed patients. Psychoneuroendocrinology 2013; 38: 863-870.
- 89. Vershinin A. Biological functions of carotenoids--diversity and evolution. Biofactors.1999; 10: 99-104.
- Moise AR, Alvarez S, Dominguez M, Alvarez R, Golczak M, et al. Activation of retinoic acid receptors by dihydroretinoids. Mol Pharmacol. 2009; 76: 1228-1237
- Foley P, Kirschbaum C. Human hypothalamus-pituitaryadrenal axis responses to acute psychosocial stress in laboratory settings. Neurosci Biobehav Rev. 2010; 35: 91-99.
- Emmert MH, Herman JP. Differential forebrain c-fos mRNA induction by ether inhalation and novelty: evidence for distinctive stress pathways. Brain Res. 1999; 845: 60-67.
- 93. Der-Avakian A, Markou A. The neurobiology of anhedonia and other reward-related deficits. Trends Neurosci. 2012; 35: 68-77.
- 94. Duncko R, Kiss A, Skultetyova I, Rusnak M, Jezova D. Corticotropin-releasing hormone mRNA levels in response to chronic mild

- stress rise in male but not in female rats while tyrosine hydroxylase mRNA levels decrease in both sexes. Psychoneuroendocrinology 2001; 26: 77-89.
- Sánchez-Sarasúa S, Moustafa S, García-Avilés A, López-Climent MF, Gómez-Cadenas A, et al. The effect of abscisic acid chronic treatment on neuroinflammatory markers and memory in a rat model of high-fat diet induced neuroinflammation. Nutr Metab. 2016; 13: 73.
- Ribes-Navarro A, Atef M, Sánchez-Sarasúa S, Beltrán-Bretones MT, Olucha-Bordonau F, et al. Abscisic Acid Supplementation Rescues High Fat Diet-Induced Alterations in Hippocampal Inflammation and IRSs Expression. Mol Neurobiol. 2019; 56: 454-464.
- 97. Jeon SH, Kim N, Ju YJ, Gee MS, Lee D, et al. Phytohormone abscisic acid improves memory impairment and reduces neuroinflammation in 5xFAD mice by upregulation of LanClike protein 2. Int J Mol Sci. 2020; 21: 8425.
- Khorasani A, Abbasnejad M, Esmaeili-Mahani S. Phytohormone abscisic acid ameliorates cognitive impairments in streptozotocin-induced rat model of Alzheimer's disease through PPARβ/δ and PKA signaling. Int J Neurosci. 2019; 129: 1053-1065.
- 99. Espinosa-Fernández V, Mañas-Ojeda A, Pacheco-Herrero M, Castro-Salazar E, Ros-Bernal F, et al. Early intervention with ABA prevents neuroinflammation and memory impairment in a triple transgenic mice model of Alzheimer's disease. Behav Brain Res. 2019; 374: 112106.
- Shabani M, Naderi R. Phytohormone abscisic acid elicits positive effects on harmaline-induced cognitive and motor disturbances in a rat model of essential tremor. Brain Behav. 2022; 12: e2564.
- 101. Sodhi RK, Singh N. All-trans retinoic acid rescues memory deficits and neuropathological changes in mouse model of streptozotocin-induced dementia of Alzheimer's type. Prog. Neuropsychopharmacol Biol Psychiatry. 2013; 40: 38-46.
- 102. Ding Y, Qiao A, Wang Z, Goodwin JS, Lee ES, et al. Retinoic acid attenuates beta-amyloid deposition and rescues memory deficits in an Alzheimer's disease transgenic mouse model. J Neurosci. 2008; 28: 11622-11634.
- 103. Bonnet E, Touyarot K, Alfos S, Pallet V, Higueret P, et al. Retinoic acid restores adult hippocampal neurogenesis and reverses spatial memory deficit in vitamin A deprived rats. PLoS ONE. 2008; 3: e3487.
- 104. Schrott L, Jackson K, Yi P, Dietz F, Johnson G, et al. Acute oral Bryostatin-1 administration improves learning deficits in the APP/PS1 transgenic mouse model of Alzheimer's disease. Curr Alzheimer Res. 2015; 12: 22-31.
- Weller J, Budson A. Current understanding of Alzheimer's disease diagnosis and treatment. F1000Res. 2018; 7: 1161.2018.
- 106. Large S, Slinger R. Grief in caregivers of persons with Alzheimer's disease and related dementia: A qualitative synthesis. Dementia. 2013; 14: 164183.
- 107. Salkovic-Petrisic M, Osmanovic-Barilar J, Knezovic A, Hoyer S, Mosetter K, et al. Long-term oral galactose treatment prevents cognitive deficits in male Wistar rats treated intracerebroventricularly with streptozotocin. Neuropharmacol. 2014; 77: 68-80.
- 108. Maixner DW, Christy D, Kong L, Viatchenko-Karpinski V, Horner KA, et al. Phytohormone abscisic acid ameliorates neuropathic pain via regulating LANCL2 protein abundance and glial activation at the spinal cord. Mol Pain. 2022; 17448069221107781.
- 109. Mollashahi M, Abbasnejad M, Esmaeili-Mahani S. Phytohormone abscisic acid elicits antinociceptive effects in rats through

- the activation of opioid and peroxisome proliferator-activated receptors  $\beta/\delta$ . Eur J Pharmacol. 2018; 832: 75-80.
- 110. Mollashahi M, Abbasnejad M, Esmaeili-Mahani S. Spinal protein kinase A and phosphorylated extracellular signal-regulated kinase signaling are involved in the antinociceptive effect of phytohormone abscisic acid in rats. Arq Neuropsiquiatr. 2020; 78: 21-27.
- Alves JM, Lin K. Neuropathic Pain: A Review of Interneuronal Disinhibition. Arch Neurosci. 2018; 5: e12290.
- 112. van Hecke O, Austin SK, Khan RA, Smith BH, Torrance N. Neuropathic pain in the general population: a systematic review of epidemiological studies. Pain. 2014; 155: 654-662.
- Liu XJ, Gingrich JR, Vargas-Caballero M, Dong YN, Sengar A, et al. Treatment of inflammatory and neuropathic pain by uncoupling Src from the NMDA receptor complex. Nat Med. 2008; 14: 1325-1332.
- Costigan M, Scholz J, Woolf CJ. Neuropathic pain: a maladaptive response of the nervous system to damage. Annu Rev Neurosci. 2009; 32: 1-32.
- 115. Ji RR, Xu ZZ, Gao YJ. Emerging targets in neuroinflammation driven chronic pain. Nat Rev Drug Discov. 2014; 13: 533-548.
- Grace PM, Hutchinson MR, Maier SF, Watkins LR. Pathological pain and the neuroimmune interface. Nat Rev Immunol. 2014; 14: 217-231.
- 117. Yan X, Weng HR. Endogenous interleukin-1β in neuropathic rats enhances glutamate release from the primary afferents in the spinal dorsal horn through coupling with presynaptic NMDA receptors. J Biol Chem. 2013; 288: 30544-30557.
- Guri AJ, Hontecillas R, Si H, Liu D, Bassaganya-Riera J. Dietary abscisic acid ameliorates glucose tolerance and obesity related inflammation in db/db mice fed high-fat diets. Clin Nutr. 2007; 26: 107-116.
- 119. Balino P, Gomez-Cadenas A, Lopez-Malo D, Romero FJ, Muriach M. Is there a role for abscisic acid, a proven antiinflammatory agent, in the treatment of ischemic retinopathies? Antioxidants. 2019; 8: 104.
- Nestler EJ. Historical review: molecular and cellular mechanisms of opiate and cocaine addiction. Trends Pharmacol Sci. 2004; 25: 210-218.
- Bishop-Bailey D, Bystrom J. Emerging roles of peroxisome proliferator-activated receptor-beta/delta in inflammation. Pharmacol Ther. 2009; 124: 141-150.
- 122. Bassaganya-Riera J, Guri AJ, Lu P, Climent M, Carbo A, et al. Abscisic acid regulates inflammation via ligand-binding domain-independent activation of peroxisome proliferator-activated receptor gamma. J Biol Chem. 2011; 286: 2504-2516.
- Zocchi E, Hontecillas R, Leber A, Einerhand A, Carbo A, et al. Abscisic Acid: A Novel Nutraceutical for Glycemic Control. Front Nutr. 2017; 4: 24.
- 124. Asmund Arnesen JA, Jacobsen IH, Dyekjær JD, Rago D, Kristensen M, et al. Production of abscisic acid in the oleaginous yeast Yarrowia lipolytica. FEMS Yeast Res. 2022; 22: foac015.
- 125. Vaidya AS, Park SY, Xing Z, Cutler SR. Synthesis and characterization of abscisic acid receptor modulators. Methods Enzymol. 2022; 671: 435-470.
- 126. Vaidya AS, Helander JDM, Peterson FC, Elzinga D, Dejonghe W, et al. Dynamic control of plant water use using designed ABA receptor agonists. Science. 2019; 366: eaaw8848.